

Loss-of-function mutations in *SCN4A* cause severe foetal hypokinesia or ‘classical’ congenital myopathy

Irina T. Zaharieva,^{1,*} Michael G. Thor,^{2,*} Emily C. Oates,^{3,4,*} Clara van Karnebeek,⁵ Glenda Hendson,⁶ Eveline Blom,⁷ Nanna Witting,⁸ Magnhild Rasmussen,^{9,10} Michael T. Gabbett,¹¹ Gianina Ravenscroft,¹² Maria Sframeli,¹ Karen Suetterlin,² Anna Sarkozy,¹ Luigi D’Argenzio,¹ Louise Hartley,¹³ Emma Matthews,² Matthew Pitt,¹⁴ John Vissing,⁸ Martin Ballegaard,¹⁵ Christian Krarup,¹⁵ Andreas Slørdahl,¹⁶ Hanne Halvorsen,¹⁷ Xin Cynthia Ye,⁵ Lin-Hua Zhang,⁵ Noline Løkken,⁸ Ulla Werlauff,¹⁸ Mena Abdelsayed,¹⁹ Mark R. Davis,²⁰ Lucy Feng,¹ Rahul Phadke,¹ Caroline A. Sewry,¹ Jennifer E. Morgan,^{1,2} Nigel G. Laing,¹² Hilary Vallance,⁵ Peter Ruben,¹⁹ Michael G. Hanna,² Suzanne Lewis,⁵ Erik-Jan Kamsteeg,²¹ Roope Männikkö² and Francesco Muntoni^{1,2}

*These authors contributed equally to this work.

See Cannon (doi:10.1093/brain/awv400) for a scientific commentary on this article.

Congenital myopathies are a clinically and genetically heterogeneous group of muscle disorders characterized by congenital or early-onset hypotonia and muscle weakness, and specific pathological features on muscle biopsy. The phenotype ranges from foetal akinesia resulting in *in utero* or neonatal mortality, to milder disorders that are not life-limiting. Over the past decade, more than 20 new congenital myopathy genes have been identified. Most encode proteins involved in muscle contraction; however, mutations in ion channel-encoding genes are increasingly being recognized as a cause of this group of disorders. *SCN4A* encodes the α -subunit of the skeletal muscle voltage-gated sodium channel (Na_v1.4). This channel is essential for the generation and propagation of the muscle action potential crucial to muscle contraction. Dominant *SCN4A* gain-of-function mutations are a well-established cause of myotonia and periodic paralysis. Using whole exome sequencing, we identified homozygous or compound heterozygous *SCN4A* mutations in a cohort of 11 individuals from six unrelated kindreds with congenital myopathy. Affected members developed *in utero*- or neonatal-onset muscle weakness of variable severity. In seven cases, severe muscle weakness resulted in death during the third trimester or shortly after birth. The remaining four cases had marked congenital or neonatal-onset hypotonia and weakness associated with mild-to-moderate facial and neck weakness, significant neonatal-onset respiratory and swallowing difficulties and childhood-onset spinal deformities. All four surviving cohort members experienced clinical improvement in the first decade of life. Muscle biopsies showed myopathic features including fibre size variability, presence of fibrofatty tissue of varying severity, without specific structural abnormalities. Electrophysiology suggested a myopathic process, without myotonia. *In vitro* functional assessment in HEK293 cells of the impact of the identified *SCN4A* mutations showed loss-of-function of the mutant Na_v1.4 channels. All, apart from one, of the mutations either caused fully non-functional channels, or resulted in a reduced channel activity. Each of the affected cases carried at least one full loss-of-function mutation. In five out of six families, a second loss-of-function mutation was present on the trans allele. These functional results provide convincing evidence for the pathogenicity of the identified mutations and suggest that different degrees of loss-of-function in mutant Na_v1.4 channels are associated with attenuation of the skeletal muscle action potential amplitude to a level insufficient to support normal muscle function. The results demonstrate that recessive loss-of-function *SCN4A* mutations should be considered in patients with a congenital myopathy.

Received August 3, 2015. Revised September 25, 2015. Accepted October 13, 2015. Advance Access publication December 22, 2015

© The Author (2015). Published by Oxford University Press on behalf of the Guarantors of Brain.

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

- 1 Dubowitz Neuromuscular Centre, UCL Institute of Child Health, London, WC1N 1EH, UK
- 2 MRC Centre for Neuromuscular Diseases, UCL Institute of Neurology, London, WC1N 3BG, UK
- 3 Institute for Neuroscience and Muscle Research, Children's Hospital at Westmead, Westmead, New South Wales, 2145, Australia
- 4 Discipline of Paediatrics and Child Health, Faculty of Medicine, The University of Sydney, Sydney, New South Wales, 2006, Australia
- 5 Department of Pediatrics, Child and Family Research Institute, Centre for Molecular Medicine and Therapeutics, University of British Columbia, 4480 Oak Street, Vancouver, B.C. V6H 3V4, Canada
- 6 Department of Pathology and Laboratory Medicine, British Columbia Children's Hospital, 4500 Oak Street, Vancouver, B.C. V6H 3N1, Canada
- 7 Maastricht University Medical Center, Maastricht, 6211 LK, Netherlands
- 8 Copenhagen Neuromuscular Center, Rigshospitalet, University of Copenhagen, DK2100 Copenhagen, Denmark
- 9 Department of Clinical Neuroscience for Children, Oslo University Hospital, 0424, Oslo, Norway
- 10 Unit for Hereditary Neuromuscular Disorders, Oslo University Hospital, 0424, Oslo, Norway
- 11 Genetic Health Queensland, Royal Brisbane & Women's Hospital & Griffith University, Brisbane, Australia
- 12 The Harry Perkins Institute of Medical Research, Centre for Medical Research, The University of Western Australia, Perth, 6009, Western Australia, Australia
- 13 Department of Child Health, University Hospital Wales, Cardiff, CF14 4XW, UK
- 14 Neurophysiology Department, Great Ormond Street Hospital for Children NHS Foundation Trust, Great Ormond Street, London WC1N 3JH, UK
- 15 Department of Clinical Neurophysiology, Rigshospitalet, University of Copenhagen, DK2100 Copenhagen, Denmark
- 16 Children's Clinic, St.Olavs hospital, Trondheim University Hospital, 7006 Trondheim, Norway
- 17 Department of Pathology, University Hospital of North Norway, 9038 Tromsø, Norway
- 18 The Danish National Rehabilitation Center for Neuromuscular Diseases, Aarhus, 8000 Denmark
- 19 Department of Biomedical Physiology and Kinesiology, Simon Fraser University, Burnaby, V5A 1S6, Canada
- 20 Department Molecular Genetics, Pathwest, QEII Medical Centre, Nedlands 6009, Western Australia, Australia
- 21 Department of Human Genetics, Radboud University Medical Center, Nijmegen, 6500HB, The Netherlands

Correspondence to: Professor Francesco Muntoni,
Institute of Child Health,
University College London,
30 Guilford Street,
London WC1N 1EH,
UK
E-mail: f.muntoni@ucl.ac.uk

Keywords: *SCN4A*; loss-of-function mutation; foetal hypokinesia; foetal akinesia; congenital myopathy

Abbreviations: ExAC = Exome Aggregation Consortium; HEK293 = human embryonic kidney 293

Introduction

Congenital myopathies are a group of clinically and genetically heterogeneous muscle disorders often characterized by specific structural abnormalities in muscle biopsies. The degree of severity varies widely from profound muscle weakness leading to *in utero* or neonatal lethality, to less severe infant- or childhood-onset weakness, which results in a much milder clinical course (Wallgren-Pettersson and Laing, 2010). Common features are prominent facial muscle weakness with or without ptosis and proximal predominant limb muscle weakness.

Congenital myopathies demonstrate a considerable degree of locus and allelic heterogeneity. Mutations in different genes can cause the same pathology, and mutations in the same gene can lead to different types of muscle pathology. This indicates that disruption of a common pathway, rather than a strict gene-pathology relationship, determines the disease pathogenesis (Sewry *et al.*, 2008; Nance *et al.*, 2012; Ravenscroft *et al.*, 2015). In the past decade, whole exome sequencing has contributed to a better understanding of the pathogenesis of

congenital myopathies, by facilitating the identification of novel disease-causing genes and by broadening our understanding of the range of phenotypes associated with known causative genes.

SCN4A encodes the α -subunit of the skeletal muscle voltage-gated sodium channel (Na_v1.4) mainly expressed in skeletal muscle. The tetrodotoxin-sensitive Na_v1.4 channel (Trimmer *et al.*, 1989; Zhou and Hoffman, 1994) is essential for the generation and propagation of the muscle action potential crucial for skeletal muscle contraction. Na_v1.4 is a large protein composed of four homologous domains (DI–DIV) each consisting of six transmembrane segments (S1–S6). Segments S5 and S6 of each domain form the single ion-conducting pore, while S1–S4 form the voltage sensing domains. The four positively charged S4 segments function as voltage sensors regulating the opening and closing of the pore (Stühmer *et al.*, 1989).

Dominant gain-of-function mutations in *SCN4A* are a well-established cause of disorders collectively termed sodium channelopathies. The range of phenotypes associated with sodium channelopathies includes episodes of

muscle stiffness (potassium-aggravated myotonia), episodes of paralytic attacks associated with reduced serum potassium levels (hypokalaemic periodic paralysis) or combination of attacks of both myotonia and periodic paralysis in the same patient (paramyotonia congenita, hyperkalaemic periodic paralysis) (Vicart *et al.*, 2005; Ryan *et al.*, 2007; Corrochano *et al.*, 2014). Attacks can be triggered by changes in temperature or ion concentration and can last from minutes to days. In myotonia, the mutated Na_v1.4 causes sarcolemmal hyperexcitability and delayed muscle relaxation after contraction, while in periodic paralysis a prolonged sodium current (hyperkalaemic periodic paralysis) or an aberrant gating pore current through Na_v1.4 (hypokalaemic periodic paralysis type 2) results in membrane depolarization and inexcitability (Suetterlin *et al.*, 2014). Neonatal hypotonia with variable feeding and respiratory difficulty has also been rarely reported in the sodium channelopathies. This is a transient phenomenon, however, which improves spontaneously (Matthews *et al.*, 2008).

Recessive loss-of-function *SCN4A* mutations are rare and have been described in only two patients with congenital myasthenic syndrome. Two heteroallelic missense *SCN4A* mutations were detected in a patient with neonatal onset of fatigable generalized weakness and recurrent attacks of respiratory and bulbar paralysis (Tsujino *et al.*, 2003). In the second patient, the onset was ~12 years of age with episodic generalized weakness lasting for hours and resulting in muscle fatigue later in life (Arnold *et al.*, 2015). A homozygous missense mutation in *SCN4A* was identified in this patient, while the heterozygous parents were asymptomatic. In both patients, the mutant Na_v1.4 channels had enhanced fast inactivation, constituting a loss-of-function effect.

Here, we describe homozygous or compound heterozygous *SCN4A* mutations in 11 individuals from six unrelated families with congenital myopathy. The affected cases had clinical phenotypes ranging from severe foetal hypokinesia resulting in early lethality to ‘classical’ congenital myopathy that improved clinically over time. Functional assessment of the mutant Na_v1.4 channels in HEK293 cells revealed full and partial loss-of-function effects constituting a novel molecular pathomechanism associated with *SCN4A* mutations.

Materials and methods

Ethical approval

This study was approved by the Health Research Authority, NRES Committee East of England – Hatfield (REC 13/EE/0398), the institutional review boards of BC Children’s Hospital and the University of British Columbia Canada (CW12-0019; H12-00067), the medical ethics committee of the Radboud University Medical Centre (2011/188), Human

Research Ethics Committee of the University of Western Australia (RA/4/1/4403).

Informed consent was obtained from all individuals included in this study or from their parents or legal guardians.

Oocytes were isolated from adult female *Xenopus laevis* following procedures that have been approved by UCL’s Biological Services Management Group and the UK Home Office.

Whole exome sequencing

Whole exome sequencing was performed in all probands from the six families included in this study. Details of the whole exome sequencing procedures are provided in the Supplementary material.

Verification of the *SCN4A* c.3145-2A>C splice site mutation

The c.3145-2A>C mutation detected in Family 4 was verified in total RNA extracted from peripheral blood cells from the proband, mother and father. Details of the procedure are provided in the Supplementary material.

Histological analysis

Muscle biopsies were taken from the probands from Families 1, 2, 3 and 4. Post-mortem samples were taken from Patient II.2 Family 4 and from all affected infants from Families 5 and 6. Histological, histochemical and immunohistochemical studies were performed as previously described (Dubowitz *et al.*, 2013).

MRI

Muscle MRI was performed in the probands from Families 1 and 2. Conventional T₁- and, in some cases, also T₂-weighted sequences were used to obtain planes from the lower limbs. Spinal MRI was undertaken in the proband from Family 1, and cerebral MRI in the probands from Families 3 and 4.

Mutagenesis

QuikChange® II (Agilent Technologies) site-directed mutagenesis kit was used to insert mutations in the human *SCN4A* clone (from Prof. Steve Cannon, University of Texas Southwestern Medical Center, Dallas, USA).

Human embryonic kidney 293 (HEK293) cells were transfected with cDNA for the sodium channel α -subunit and green fluorescent protein (GFP) using Lipofectamine™ 2000 (Life Technologies).

Whole-cell electrophysiology

Electrophysiology methods are detailed in the Supplementary material. The extracellular solution for HEK293 recordings contained (in mM): 145 NaCl, 4 KCl, 1.8 CaCl₂, 1 MgCl₂, and 10 HEPES (pH = 7.35) and the intracellular pipette solution contained (in mM): 145 CsCl, 5 NaCl, 10 EGTA, 10 HEPES (pH = 7.4). The calculated liquid junction potential is –4.4 mV, which was not corrected for. Holding potential was

–80 mV. Leak and capacitance subtraction was performed online using a $-P/4$ procedure.

The voltage protocols are described in the legends and in the Supplementary material. Functional characterization in *X. laevis* oocytes is described in the Supplementary material.

Statistical analysis

Data analysis, presentation and curve-fitting protocols are described in the Supplementary material. Data are expressed as mean \pm standard error of the mean (SEM). Statistical significance was analysed using Student's *t*-test and accepted at $P < 0.05$.

Results

Identification of SCN4A mutations in the congenital myopathy patients

Whole exome sequencing was carried out in the proband from Family 1 as part of the European Commission funded NeurOmics project. Following a filtering procedure (de Ligt *et al.*, 2012) and assuming a recessive mode of inheritance, we were left with eight high quality variants (Supplementary Table 3). Only the two heterozygous variants in *SCN4A* were not present in the Exome Aggregation Consortium (ExAC) database and were predicted to be pathogenic and thus were considered as the disease-causing mutations. Segregation analysis showed that the compound heterozygous *SCN4A* c.311G>A (p.R104H) and c.3403C>T (p.R1135C) mutations detected in the proband had been inherited from the unaffected mother and father, respectively.

Through an international collaborative effort we identified 10 additional cases with congenital myopathy from five unrelated families (Fig. 1). The affected cases carried homozygous or compound heterozygous *SCN4A* variants that were predicted to be pathogenic. All 11 *SCN4A* mutations identified in the cohort members are described in Fig. 2. The *SCN4A* mutations were validated by Sanger sequencing and segregation consistent with an autosomal recessive pattern of inheritance was confirmed in all six families. The location of the mutations relative to the domains of the Na_v1.4 protein is presented in Fig. 2A.

Consistent with mutations responsible for a rare early-onset recessive disorder, eight of the variants had not been reported previously in ExAC, and three mutations (p.Q470X, p.H1782Qfs65 and p.D1069N) were present at very low frequency (<0.00001) (Fig. 2B).

Of the 11 *SCN4A* mutations identified, nine had not been previously reported, p.R1135C has been described in a homozygous state in a patient with hypokalaemic periodic paralysis (Groome *et al.*, 2014) and p.R225W was reported in a heterozygous state in a patient with mild non-dystrophic myotonia (Lee *et al.*, 2009).

Clinical features of cohort members

We detected a continuum of clinical severity ranging from severe foetal hypokinesia resulting in early lethality in seven cases, to a milder congenital myopathy phenotype in four surviving individuals. Phenotypic features of the cohort are summarized in Table 1 and clinical data are detailed in Supplementary Table 1. Clinical and muscle MRI images are shown in Fig. 3. The most salient clinical features are described below.

Clinical features of severely affected cohort members with foetal hypokinesia

In utero history

The seven most severely affected cohort members developed marked foetal hypokinesia from 19 to 32 weeks gestation. All seven also developed *in utero*-onset limb contractures and/or talipes, which were first detected at 19 to 31 weeks. Talipes was the most common limb deformity (seen on ultrasound in 5/7 severely affected infants; first detected between 19 to 27 weeks). Additional lower limb joint contractures were noted prior to delivery in five of the infants. *In utero*-onset upper limb and/or finger contractures were detected on ultrasound in four cases, sometimes in combination with lower limb contractures (three cases). Almost all severely affected cohort members also developed features suggestive of hydrops (present in 6/7; evident from 20 to 31 weeks). The mothers of all seven severely affected cohort members developed polyhydramnios (onset 20 to 31 weeks), which typically developed in parallel with hypokinesia and foetal limb contractures. Decompression procedures to reduce liquor volume were required in two cases.

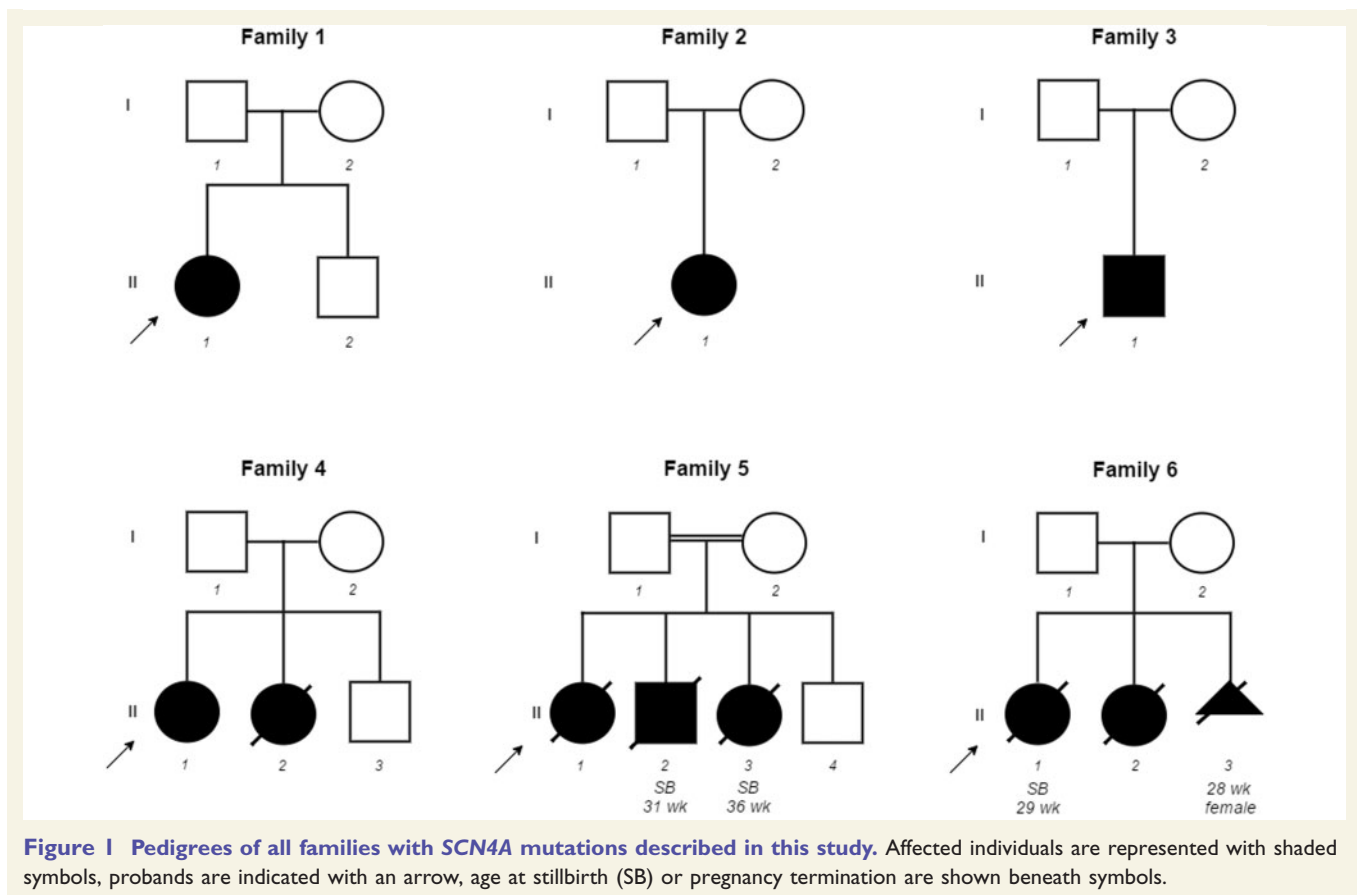
Prematurity and early death

One severely affected infant was born at term but died at 5 h of age from respiratory complications. The other six severely affected infants died at or before 36 weeks gestation: two infants died spontaneously prior to delivery (at 29 and 31 weeks gestation), one died during delivery (at 36 weeks), two died within 8 h of their premature delivery due to severe respiratory insufficiency (at 32 and 33 weeks), and one pregnancy was terminated at 28 weeks gestation due to recurrence of features seen in two severely affected pregnancies.

All seven cohort members who did not survive beyond the first day of life had at least one sign of *in utero* involvement (e.g. hypokinesia, limb contractures or talipes) by 32 weeks gestation, and in 4/7 cases at least one abnormality was evident by 20 weeks.

External examination and autopsy findings

External examination was undertaken in all seven infants who died prematurely or at term. Six also underwent autopsy. Marked muscle hypoplasia was present in all seven. Hydrops was confirmed in six cases. External and autopsy examination revealed features associated with hydrops (e.g. pericardial and pleural effusions, ascites and/or hepatomegaly), in addition to the features associated with severe foetal



hypokinesia (a small thorax, pulmonary hypoplasia, limb contractures and fractures). In addition to talipes (present in 6/7 cases), four had lower limb contractures; typically bilateral hip flexion contractures (3/4) with or without additional external rotation, and knee extension contractures (4/4). Five had upper limb contractures (finger flexion in 4/7, wrist 1/7, elbow flexion 2/7, shoulder flexion 1/7). In general, lower limb contractures were more marked than upper limb contractures. Limb fractures were present in 2/7 cases (left humerus and right mid-femur). Thin gracile ribs and/or thin long bones were noted in at least 2/7 cases. Several infants had additional facial features that were not shared by other family members. This included plagiocephaly (1/7), frontal bossing (2/7), deep set eyes (2/7 cases), downward-sloping lateral orbital ridges (1/7), a flattened nose (3/7), ear abnormalities (3/7), retrognathia and/or micrognathia (3/7), and/or a small triangular or tented mouth (2/7). A subset of these (e.g. multiple joint contractures, hydrops/oedema, pulmonary hypoplasia, and retro-micrognathia) are well-known features of foetal akinesia deformation sequence (Ravenscroft *et al.*, 2011).

Clinical features in surviving cohort members with 'classical' congenital myopathy

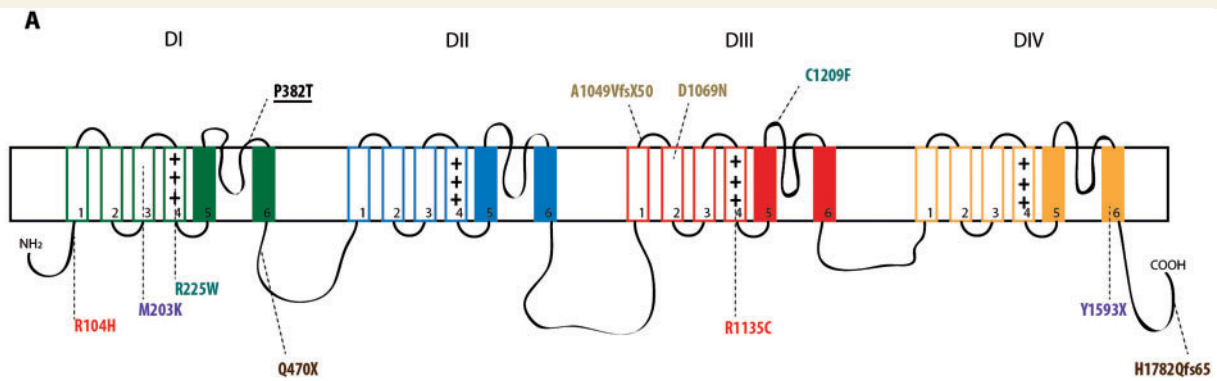
The four surviving cohort members are currently aged 2.5, 8, 14 and 35 years, and all have features consistent with 'classical' congenital myopathy.

In utero and birth history

Pregnancy records were available for three of the four surviving cohort members, with limited data available for the fourth. *In utero* features were generally less marked than in severe foetal hypokinesia cases; however, at least one abnormality (hypokinesia, polyhydramnios or talipes) was present in three of the four cases. Foetal hypokinesia was noted during the pregnancies of two of the four surviving cohort members (onset at 20 and 31 weeks, respectively), but appears to have been less marked than in more severely affected cohort members. Polyhydramnios complicated the pregnancies of 2/4 surviving cohort members and was first noted slightly later (31 to 40 weeks). None of the surviving cohort members had limb contractures or talipes detected on prenatal ultrasound, or a history of hydrops. All four were born at or close to term. Breech presentation, or transverse lie, complicated delivery of three of these four affected individuals. One required resuscitation following delivery.

Congenital features

All four surviving cohort members had moderate-to-severe hypotonia and facial, neck, axial and limb weakness, which was either present at birth (3/4) or developed within the first few days of life (as was the case for the proband from Family 3). A weak cry, a generalized reduction in muscle bulk, and reduced or absent limb reflexes were also each



B

Family (member)	Family 1 (II.1)		Family 2 (II.1)		Family 3 (II.1)		Family 4 (II.1, II.2)		Family 5 (II.1, II.2, II.3)	Family 6 (II.1, II.2, II.3)	
SCN4A Mutation	c.311G>A	c.3403C>T	c.673C>T	c.3628G>T	c.1408C>T	c.5345dup	c.3205G>A	c.3145-2A>C	<u>c.1144C>A</u> (homozygous)	c.608T>A	c.4779C>A
Amino Acid change	p.R104H	p.R1135C	p.R225W	p.C1209F	p.Q470X	p.H1782Qfs65	p.D1069N	p.A1049VfsX50	p.P382T	p.M203K	p.Y1593X
Mutation type	MS	MS	MS	MS	NS	FS	MS	ESS	MS	MS	NS
ExAC Allele Frequency	-	-	-	-	0.000008	0.000008	0.000017	-	-	-	-
Reported in Sodium Channelopathy	-	HypoPP	NDM	-	-	-	-	-	-	-	-

Figure 2 SCN4A mutations in congenital myopathy patients. Location of the mutations mapped onto a secondary structure of Na_v1.4 channel (A). Compound heterozygous mutations identified in the affected individuals from one family are presented with the same colour. The homozygous mutation in Family 5 is shown underlined. Position, amino acid change, mutation type, frequency in ExAC database and association with sodium channelopathy of the reported mutations (B). Full loss-of-function mutations are presented in red. MS = missense mutation; NS = nonsense mutation; FS = frameshift mutation; ESS = essential splice site mutation; HypoPP = hypokalaemic periodic paralysis; NDM = non-dystrophic myotonia; - = not available.

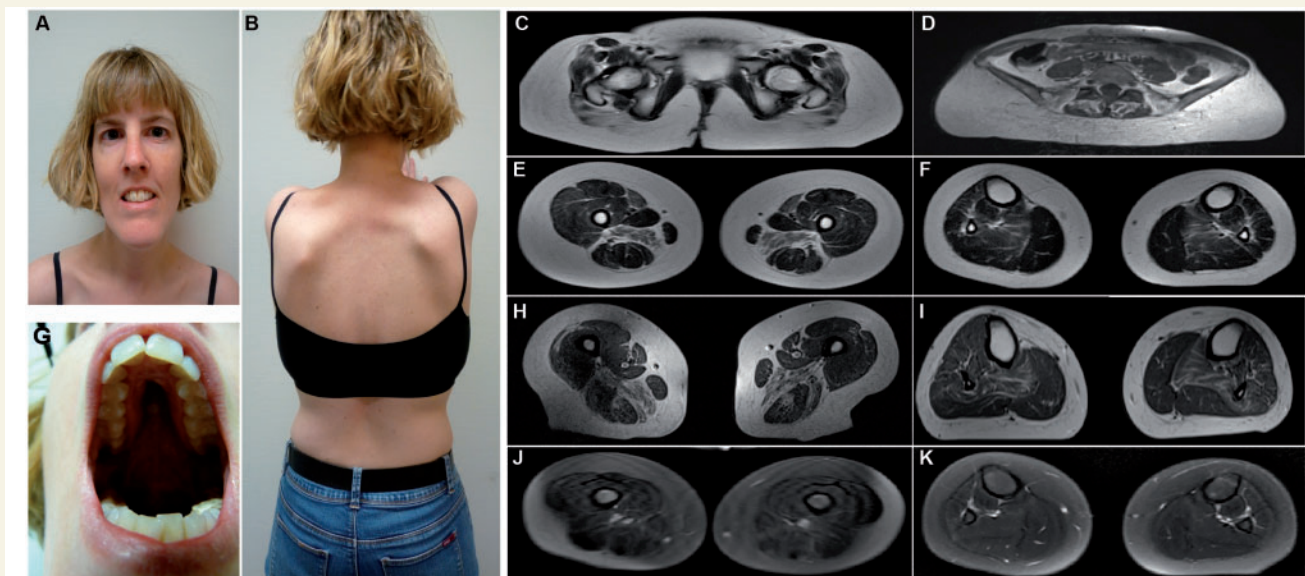


Figure 3 Clinical features in SCN4A congenital myopathy patients. Clinical images of the proband from Family 2 show mild facial weakness, elongated face (A), high arched palate (G) and bilateral scapular winging (B). T₁-weighted muscle MRI images in the affected case from Family 1 (at age 6 years) (C, E and F) and in the proband from Family 2 (at age 35 years) (D, H and I) showed severe involvement of the gluteal muscles (C and D), bilateral, symmetric involvement of sartorius and adductor magnus in the upper leg (E and H) and involvement of soleus in the lower leg (F and I). T₂-weighted muscle MRI images in the affected case from Family 1 demonstrated no oedema (J and K).

Table 1 Clinical characteristics of congenital myopathy patients with homozygous or compound heterozygous *SCN4A* mutations

Family Age Gender	<i>In utero</i> and congenital features in surviving cohort members	Muscle involvement and progression over time in surviving cohort members	Additional features in surviving cohort members
Family 1 14 y Female	<p>Pregnancy/delivery: Reduced movements Breech presentation</p> <p>Congenital features/early support: Moderate to severe congenital hypotonia Weak cry Thin muscle build Talipes Tube fed for first 12 days: Ongoing difficulties with suck during infancy but able to bottle feed</p>	<p>Pattern of muscle involvement: Mild facial weakness, high arched palate, elongated face Neck flexion and axial weakness Limb weakness: PUL + +, DUL +, PLL + + +, DLL + + Weakest muscle groups: neck flexors, axial muscles, hip extensors and abductors Generalized muscle atrophy most marked in shoulders</p> <p>Progression: Oromotor difficulties resolved by 2 y Hip, knee and tendo Achilles contractures from 2 y Delayed motor milestones: first walked independently 2 y 9 m Able to jump and run (slowly) by 3–4 y In childhood: frequent falls, positive Gowers' sign Ongoing improvements in strength and motor skills over time</p> <p>Current status: Still ambulant (500 m) with slow waddling gait Uses splints (since age 4 y), K-walker and manual wheelchair for longer distances</p>	<p>Mild ophthalmoplegia (upgaze weakness) noted 3 y 9 m Large asymmetrical dolichocephalic head shape, frontal bossing, micrognathia Scoliosis and spinal rigidity first noted age 2 y: spinal fusion for scoliosis aged 12 y BIPAP required from 6 y: most recent FVC 44% (13 y 10 m) Pes planus noted in childhood Height initially 50th percentile: fell to 2–10th percentile by 13 y 10 m (in part due to scoliosis) Hypermobility Mild asymmetrical pectus excavatum Short-lived episodic weakness during febrile illnesses, post-exercise, and on hot days since 12 y Activity-limiting increase in fatigability from 13 y: fatigues quickly with walking and writing Improved strength endurance with regular oral salbutamol</p>
Family 2 35 y Female	<p>Pregnancy/delivery:^L Breech presentation</p> <p>Congenital and neonatal features/early support:^L Moderate to severe congenital hypotonia Non-invasive respiratory support needed soon after birth^L</p>	<p>Pattern of muscle involvement: Mild facial weakness, high arched palate, elongated face Neck flexion and axial weakness Limb weakness: PUL 4/5, DUL 5/5, PLL 3/5, DLL 5-/5 Weakest muscle groups: neck flexors, axial muscles, hip extensors and abductors Generalized muscle atrophy most marked in sternocleidomastoids</p> <p>Progression: Delayed motor milestones^L Positive Gowers' sign Ongoing improvements in strength and motor skills over time in early life. Deterioration in motor abilities from 30 y</p> <p>Current status: Still ambulant (500 m) with slow waddling gait</p> <p>Pattern of muscle involvement: Moderate facial weakness, high arched palate Neck weakness</p>	<p>Recurrent cyanotic spells during infancy: cause not known: ceased after 2 y Generalized ophthalmoplegia Hypernasal voice Scapular winging Fatigues quickly when walking FVC 83% at 30 y Multiple dental procedures to manage dental crowding</p>
Family 3 2.5 y Male	<p>Pregnancy/delivery: Polyhydramnios</p>	<p>Current status: Still ambulant (500 m) with slow waddling gait</p> <p>Pattern of muscle involvement: Moderate facial weakness, high arched palate Neck weakness</p>	<p>Urachus excision at 3 m Kyphosis first noted 22 m</p>

(continued)

Table 1 Continued

Family Age Gender	In utero and congenital features in surviving cohort members	Muscle involvement and progression over time in surviving cohort members	Additional features in surviving cohort members
Family 4 8 y Female	<p>Congenital and neonatal features/early support: Moderate to severe neonatal hypotonia (increase noted during first 3 days) Cried infrequently Significant ongoing hypotonia, feeding and respiratory difficulties during early infancy CPAP from day 3 to day 7 of life: recommenced at 7 weeks: BiPAP from 4 m: predominantly during sleep: ongoing NG fed from 1–2 weeks: PEG fed from 3 m: ongoing but trial of oral feeding planned Tricuspid insufficiency noted at 3 days: no long term sequelae</p> <p>Pregnancy/delivery: Foetal hypokinesia from 31/40, polyhydramnios from 31–32/40 Transverse lie Resuscitation at birth</p> <p>Congenital features/early support: Moderate to severe congenital hypotonia Weak cry Thin muscle build Hip contractures: resolved with passive stretching exercises Intubated from birth until 13 m: BiPAP at night and during daytime sleeps until 2 y NG/PEG fed from birth until 4 y</p>	<p>Limb weakness: PUL + + +, DUL + + +, PLL + + +, DLL + + + Weakest muscle groups: neck and interscapular muscles Generalized muscle atrophy</p> <p>Progression: Delayed motor milestones: first sat independently before 2 y: not yet walking. Ongoing improvements in strength and motor skills over time Additional expressive language and fine motor delay</p> <p>Current status: Able to take a few steps with frame, uses a wheelchair</p> <p>Pattern of muscle involvement: Mild facial weakness, high arched palate, elongated face Neck and axial weakness Limb weakness: PUL + + +, DUL + + +, PLL + + +, DLL + + + Weakest muscle groups: axial muscles Generalized muscle atrophy most marked axial muscles</p> <p>Progression: Delayed motor milestones: first walked independently 18 m Able to jump and run (slowly) by 3 y Ongoing improvements in strength and motor skills over time Additional persisting oromotor weakness, expressive language and fine motor delay</p> <p>Current status: Still ambulant: awkward, broad-based, uncoordinated gait</p>	<p>Dolichocephalic head shape, synophrys, proptosis, deep set eyes, large down-slanting palpebral fissures, broad nasal root and bridge Upper thoracic scoliosis first noted 3 y Generalized hypermobility with foot valgus deformity and pectus excavatum (Marfanoid habitus) Unilateral scapular winging Fatigues quickly when walking and writing Persisting oromotor weakness and expressive language delay</p>
Family 5 (11.1) 33/40 minutes post delivery Female	<p>Hypokinesia from 32/40 Finger flexion contractures Polyhydramnios from 31–32/40: Two liquor reduction procedures Breech presentation: no limb movement or respiratory effort at birth Marked generalized muscle hypoplasia Hypokinesia, hydrops and polyhydramnios from 20/40 Upper and lower limb contractures from 20/40, including talipes Induced after SROM Marked generalized muscle hypoplasia</p>	<p>Upper limbs: Bilateral finger flexion contractures No lower limb contractures or foot deformities</p> <p>Face: deep set eyes, flattened nose/facies, abnormal outer ears, tented mouth, retrognathia/micrognathia, frontal bossing Trunk: small thorax Upper limbs: elbow flexion contractures, wrist contractures (eversion), finger contractures Lower limbs: hip flexion contractures, knees in extension Feet: bilateral talipes EV</p>	<p>Autopsy not performed</p> <p>Lungs: pulmonary hypoplasia Brain: mild periventricular leukoencephalopathy: likely secondary to ischaemia and chorioamnionitis</p>

(continued)

Table 1 Continued

Family (member) Gestation at time of death Gender	In utero and congenital features in deceased cohort members	External examination findings in deceased cohort members	Internal examination findings in deceased cohort members
Family 5 (11.2) 31/40 ^{prior to delivery} Male	Talipes from 19/40; other upper and lower limb contractures from 20/40 Hypokinesia from 20/40 Hydrops from 22/40 Polyhydramnios from 26/40 Spontaneous intrauterine death Marked generalized muscle hypoplasia and diffuse/extensive hydrops/oedema	Face: large head, flattened nose, retrognathia/micrognathia Upper limbs: elbow contractures, finger contractures (milder than in Patient II.1) Lower limbs: knee contractures (milder than in Patient II.1) Feet: bilateral foot eversion (fixed)	Lungs: pulmonary hypoplasia
Family 5 (11.3) 36/40 ^{during delivery} Female	Hypokinesia and polyhydramnios from 24/40 Lower limb contractures evident from 25/40 Talipes evident from 24/40 Hydrops from 25/40 Liquor reduction procedure at 28/40 No spontaneous respiration at birth: resuscitation unsuccessful Marked generalized muscle hypoplasia and extensive hydrops/oedema	Face: deep set and small eyes, flattened nose, small nose, low-set ears Upper limbs: bilateral 2nd and 3rd finger flexion contractures Lower limbs: hip flexion contractures, knees in extension Feet: bilateral talipes EV	Fracture: left humerus Lungs: pulmonary hypoplasia Liver: hepatomegaly
Family 6 (11.1) 29/40 ^{prior to delivery} Female	Polyhydramnios from 20/40 Talipes from 24/40 Hydrops noted at birth (29/40) Spontaneous OL Foetal heart beat lost during delivery: resuscitation unsuccessful Marked generalized muscle hypoplasia and extensive hydrops/oedema	Face: fontanelles small for gestation, low-set ears Upper and lower limbs: restricted ROM in shoulders, elbows, hips and knees, talipes Fingers and toes: tapering fingers; hypoplastic toenails Limb lengths: mildly reduced for gestation	Fracture: right mid-femur Other bone features: thin gracile 'ribbon-like' ribs, long thin bones Lungs: pulmonary hypoplasia (severe) Pericardial effusion, pleural effusion and ascites Excess subdural fluid Lungs: pulmonary hypoplasia (severe) Pleural effusion and ascites Brain: possible hypoplastic pyramidal tracts
Family 6 (11.2) 32/40 ^{8 hours post delivery} Female	Hypokinesia from 27/40 Lower limb contractures and talipes from 27/40 Upper limb contractures from 31/40 Polyhydramnios from second trimester Hydrops from 31/40 Emergency caesarean section Marked generalized muscle hypoplasia and mild hydrops/oedema: most marked around head and neck	Face: mild frontal bossing, downward sloping lateral orbital ridges, small nose, small triangular mouth with narrow vermilion borders, straight lower lip, short philtrum, short neck, retrognathia/micrognathia Trunk: small/narrow trunk including thorax Upper and lower limbs: clenched fingers, and restricted ROM in large joints Feet: bilateral talipes EV	Lungs: pulmonary hypoplasia Pleural effusion and ascites Brain: possible hypoplastic pyramidal tracts
Family 6 (11.3) 28/40 ^{pregnancy terminated} Female	Hypokinesia from 19/40 No upper limb contractures Lower limb contractures evident from 19/40 No talipes Polyhydramnios from 28/40 Hydrops from 28/40	Marked generalized muscle hypoplasia and mild hydrops/oedema Upper limbs: left shoulder flexion contracture, fingers held in flexion but able to be extended Lower limbs: hip flexion contractures, knees in extension Feet: bilateral talipes (feet and toes plantar-flexed)	Lungs: pulmonary hypoplasia

'X'/40 = 'X' weeks gestation. For example 31/40 = 31 weeks gestation; BiPAP = bilevel positive airway pressure; CPAP = continuous positive airways pressure; d = days; DLL = distal lower limb; FVC = forced vital capacity; † = limited data available; NG = nasogastric; PEG = percutaneous endoscopic gastrostomy; OL = onset of labour; PLL = proximal lower limb; PUL = proximal upper limb; DUL = distal upper limb; ROM = range of motion; SROM = spontaneous rupture of membranes; EV = equinovarus; y = years; m = months.

noted in two or more surviving infants. Talipes was present in only one of the four, and this was not detected until delivery. Another child had mild bilateral hip contractures first noted at birth, which resolved with stretching exercises.

Bulbar involvement

Swallowing difficulties, severe enough to warrant supplemental tube and/or parenteral gastrostomy (PEG) feeding, were present from birth in 3/4 surviving cohort members. Reliance on supplemental feeding typically declined over time, and ceased during infancy or early childhood. The two youngest cohort members (now aged 2.5 and 8 years) have evidence of persisting oromotor weakness and expressive language delay suggestive of ongoing bulbar involvement, however both have experienced improvement in their feeding and language status, over time.

Pattern of muscle involvement and contractures

All surviving cohort members had mild-to-moderate upper and lower limb weakness, which was typically more marked proximally (3/4 cases). Hip extension and abduction were the weakest manoeuvres in 2/4 cases. Neck weakness was universal (4/4), and typically more pronounced than limb weakness. Neck flexion was usually weaker than extension. Shoulder, neck and truncal muscle atrophy was often more marked than limb muscle atrophy. Muscle hypertrophy, muscle pain and fasciculations were not present in any cohort member.

Postnatal-onset lower limb contractures were noted in only 1/4 surviving cohort member. Abnormal foot positioning was noted in 2/4 (pes planus in 1/4, hindfoot valgus/forefoot pronation deformities in 1/4).

Progression in axial weakness was documented in at least one surviving cohort member. This resulted in the development of respiratory insufficiency severe enough to warrant institution of non-invasive nocturnal ventilation at age 6 years. Two of the four surviving cohort members needed non-invasive respiratory support at or soon after birth, and a third required invasive ventilation. The duration of support varied; however, two were weaned from respiratory intervention by the age of 2 years. The third remains reliant on nocturnal BiPAP (bilevel positive airway pressure) at age 2.5.

Postnatal-onset scoliosis, kyphosis and spinal rigidity each were also reported in at least one additional case.

Facial and eye involvement

Mild-to-moderate facial weakness and myopathic facial features including an elongated face and/or a high arched palate were present in all four cases (Fig. 3). Additional dysmorphisms noted in at least 1/4 included dolichocephaly, synophrys, proptosis, large down-slanting palpebral fissures, and a broad nasal root and bridge. A subset of the dysmorphic facial features noted in cohort members with severe foetal hypokinesia, including deep set eyes, frontal bossing and micrognathia were present in at least one surviving individual. Mild weakness of up-gaze developed in

one child at age 3 years. Generalized ophthalmoplegia was noted in a second affected individual. Ptosis was not present in any cohort member.

Functional abilities

Delay in early motor milestones was universal. Age when independent ambulation was first achieved ranged from 18 months to 2 years 9 months. At least two cohort members were eventually able to run, albeit slowly and with difficulty. Gait was typically broad-based, slow, and waddling. Two of four used assistive devices (splints and/or frame), and 2/4 used a wheelchair. At least two cohort members had additional fine motor delay. All four experienced clearly documented improvements in strength and motor skills over time. All three that achieved independent ambulation remain able to walk. The youngest (2.5 years old) member is making good progress towards walking independently.

Additional features

Other features noted in two or more affected individuals included chest wall deformities (2/4), generalized joint hypermobility (at least 2/4), unilateral or bilateral scapular winging (one case each) (Fig. 3). One surviving cohort member had tricuspid insufficiency at birth, which resolved with time. No cardiac defects were present in the other three individuals, and none have developed additional cardiac complications over time.

Intermittent exacerbations in muscle weakness and increased fatigability

Marked fatigability with walking and/or writing was a feature in at least 3/4 of the surviving cohort members.

The proband from Family 1 (carrying heterozygous p.R1135C), who is now 14 years of age, began to experience exercise- and illness-associated exacerbations in her level of muscle weakness from the age of 12, and an activity-limiting increase in her fatigability from age 13. These episodes can last a few minutes and while they could be considered reminiscent of periodic paralysis, her repetitive stimulation and single fibre EMG results were normal, with no evidence of alteration of the amplitude or the total area of the compound muscle action potential during exercise test performed after cooling or after a long exercise test.

The oldest cohort member (now aged 35) has experienced a deterioration in her strength and motor abilities, and increase in fatigability over the past 5 years. She also has an abnormal repetitive stimulation test result suggestive of an additional neuromuscular junction abnormality (Table 2), which may be contributing to her fatigability. One of her mutations was reported previously in a family with autosomal dominant non-dystrophic myotonia (Lee *et al.*, 2009); however, myotonia was notably absent in this and all other cohort members.

Status of confirmed carriers

Segregation studies confirmed that all 12 parents, and one unaffected sibling (from Family 4) were carriers of one

Table 2 Investigations performed in cohort members with ‘classical’ congenital myopathy

Cohort Country of origin	Family F1 UK	Family F2 Denmark	Family F3 Norway	Family F4 Canada
Creatine kinase (CK) level Normal/Abnormal (level)	Normal (21 U/l)	Normal (118 U/l)	Normal (62 U/l)	Normal (30 U/l)
Neurophysiology results EMG	At age 14 years and 8 months: Normal Short exercise test with cooling: normal Long exercise test: no significant alteration in amplitude or area CMAP amplitude (abductor digiti minimi) showed values between 4.3 and 6.4 mV. Area was between 50.6 and 55.1 ms*mv At age 5 years 2 months: Normal single fibre EMG result At age of 4 years: No decrement at 3 Hz At 14 years and 8 months: Normal	At age 35 years: Increased MUP in one muscle, normal in all other but generalized abnormal responses probably due to membrane dysfunction CMAP amplitude (ulnaris): from wrist to adm the peak-to-peak value was 5.2 mV Insufficient signal At age 35 years: No decrement at 3 Hz, 60% decrement at 10 Hz At age 35 years: Reduced motor amplitudes probably due to abnormal membrane function	At age 1 year: Not significant pathology at 1 year (vastus lateralis duration times in lower normal range) CMAP amplitude (ulnaris) showed value of 5.4 mV	Undertaken in Patient II.1: At 14 months: widespread myopathic changes in all limbs: normal motor units admixed with small amplitude polyphasic units: no spontaneous activity present, or evidence of myotonia At 6 years: no myopathic changes: all results within normal limits Note: resolution of EMG abnormalities coincided with clinical improvement
Single fibre EMG				
Repetitive nerve stimulation			At 6 weeks of age: Normal	
Nerve conduction velocity			At 6 weeks of age: Normal At 1 year of age: Normal	Undertaken in Patient II.1: At 45 days: loss of sensory nerve amplitude: motor evoked responses were normal At 14 months: still abnormal: the amplitudes, conduction velocities and distal latencies in the right upper and lower limb sensory and motor nerves were normal, as were F responses. Right sural and right radial sensory responses were still, however, absent At 6 years: Normal Note: early abnormal results thought to be due to a combination of technical and maturational effects
Muscle MRI				
Pelvis/upper leg: muscles significantly involved	At age 6 years 5 months: Severe involvement of glutei, marked bilateral involvement of sartorius and adductor magnus Soleus	At age 35 years: Severe involvement of glutei, marked bilateral involvement of sartorius and adductor magnus Soleus		
Lower leg: muscles significantly involved		No oedema		
Other relevant findings	No oedema			
Other relevant investigations/results				
Cranial MRI			At 15 months of age: Normal	Undertaken in Patient II.1: At 14 days: Normal
Spinal MRI	At age 9 years 8 months: Scoliosis present. Normal intraspinal appearances. Erector spinae muscles atrophic with fatty replacement.			
Mitochondrial studies				Respiratory chain enzyme defects noted (significant reduction in activity of all complexes including the marker enzyme, citrate synthase); all thought to be secondary changes

- = not done; MUP = motor unit potential.

SCN4A mutation. Eight parents (from Families 1, 4, 5 and 6) and the sibling carrier were examined by an experienced neurologist and/or clinical geneticist and showed no evidence of myopathy, myotonia or other abnormal neurological features. This included the paternal carrier of p.R1135C mutation (previously reported in an unusual recessive form of hypokalaemic periodic paralysis). The remaining four parents were unavailable for formal examination but by report, had no neurological symptoms or signs. This included the maternal carrier of the p.R225W mutation (previously reported in dominant non-dystrophic myotonia).

Investigations

Investigations performed in the cohort members are summarized in Table 2. The creatine kinase level was normal (21–118 U/l) in all four surviving cohort members.

Neurophysiology

Neurophysiology testing was undertaken in all four cohort members who survived infancy. EMG findings were normal in 2/4. Mild abnormalities were present in 1/4. In Patient II.1 from Family 4, there were widespread myopathic changes in all four limbs at age 14 months, however on repeat testing at age 6 years all abnormalities had normalized, in parallel with a gradual improvement in strength and motor abilities. Single fibre EMG was normal in one cohort member. Repetitive stimulation testing was undertaken in two cohort members. Neither had a positive decrement at 3 Hz. In the oldest cohort member there was, however, a 60% decrement at 10 Hz.

Nerve conduction velocities were normal in 2/4 members. Testing undertaken in the oldest cohort member, at age 35 years, showed reduced motor amplitudes. The fourth surviving cohort member had sensory conduction abnormalities at age 45 days and 14 months, but these had normalized by 6 years of age.

Muscle, spinal and cerebral magnetic resonance imaging

Lower limb muscle MRI was undertaken in Patients II.1 Family 1 (at age 6 years) and II.1 Family 2 (at age 35 years) (Fig. 3). Both patients had severe involvement of the glutei, marked bilateral, symmetric involvement of sartorius, adductor magnus and soleus. Spinal MRI was undertaken in one cohort member and showed a scoliosis and erector spinae muscle atrophy but a normal spinal cord.

Cerebral MRI was performed in two cohort members (one at 14 days, the other at 15 months of age) and was normal in both.

Muscle pathology

Post-mortem muscle samples were taken from all foetal hypokinesia affected cohort members: Patient II.2 Family 4 (gestational age 40/40); Family 5 Patient II.1 (33/40), Patient II.2 (31/40), and Patient II.3 (36/40); and Family 6 Patient II.1 (29/40), Patient II.2 (32/40), and Patient II.3 (28/40). Muscle histopathology findings included abnormal fibre size variation with a mixture of small and

large fibres across fascicles without overt grouping. Presence of fibrofatty tissue, which was particularly severe in Patient II.1 Family 6, was noted in all cases (Fig. 4A–C). Necrosis and regeneration were not seen. Rods, other inclusions, cores or other structural abnormalities were absent (Fig. 4).

Quadriceps muscle biopsies taken from the congenital myopathy cohort members showed milder myopathic features (Fig. 4). A combination of slow fibre hypotrophy with normal to larger sized fast fibres and mild slow predominance was seen in Patient II.1 Family 1. No overt structural abnormalities or nemaline rods were present (Fig. 4).

Where fibre typing data were available, muscle samples in the foetal hypokinesia cohort showed fast predominance, probably related to immaturity, while mild to marked slow predominance was observed in the biopsies from the congenital myopathy cohort.

SCN4A nonsense, essential splice site and frameshift mutations alter essential functional domains of the Na_v1.4 channel

The truncating mutations p.Q470X and p.Y1593X would delete 1366 and 240 amino acids, respectively from the C-terminal end of the Na_v1.4 protein. They would result in nonsense-mediated decay of the SCN4A transcript or if translated in non-functional channels.

To investigate the effect of the c.3145-2A>C mutation on SCN4A mRNA splicing, we performed reverse transcription PCR in peripheral blood cells from the proband, mother and father from Family 4. Using primers flanking exon 15–17, two distinct fragments were amplified. A smaller fragment was obtained in all three samples while a larger fragment was only present in the index case and the father. Sanger sequencing demonstrated the smaller to be a 424 bp fragment corresponding to the wild-type SCN4A transcript of exons 15–17. The larger 971 bp fragment contained exons 15–17 and an additional 547 bp of intron 16 (Supplementary Fig. 1). The retained intron 16 sequence introduces a stop codon in the SCN4A mRNA coding sequence at amino acid position 1099 (p.A1049VfsX50) that would generate a truncated Na_v1.4 protein. The deletion of 738 C-terminal amino acids is predicted to induce complete loss of Na_v1.4 channel function.

The frameshift p.H1782Qfs65 located at the channel C-terminus caused by c.5345 duplication generates a protein that is nine amino acids longer than the wild-type. The mutated channels would have C-terminal sequence that is completely altered compared to wild-type channels. The effect of C-terminus frameshift mutations is largely unknown and therefore p.H1782Qfs65 was included in the functional analyses.

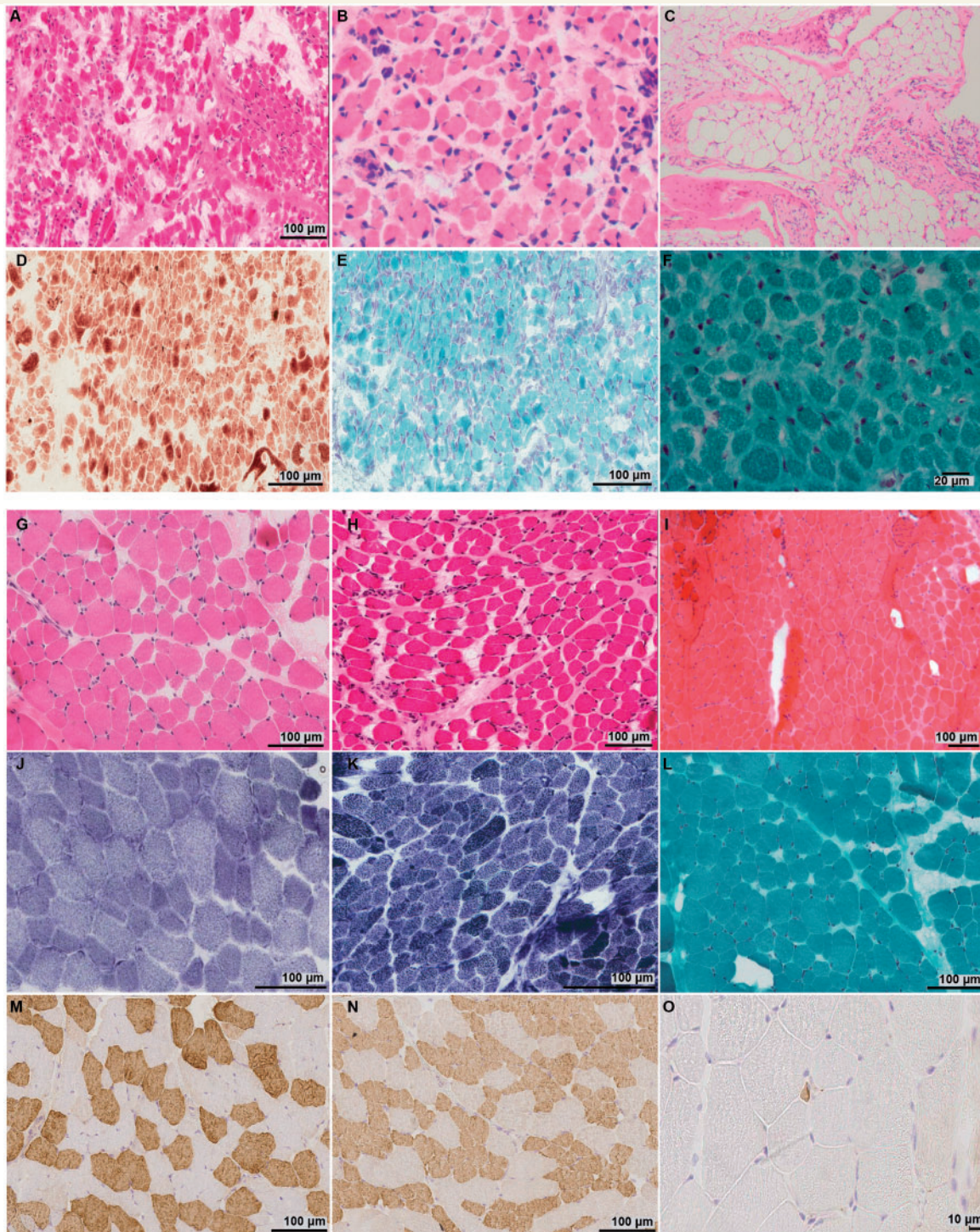


Figure 4 Muscle pathology in the foetal hypokinesia and 'classical' congenital myopathy affected individuals. Haematoxylin and eosin staining performed in the muscle samples taken from the foetal hypokinesia cases showed abnormal fibre size variation (A–C). Presence of fibrofatty tissue was noted in the muscle samples taken from affected foetuses II.2 from Family 4 and II.2 from Family 6 (A and B). Marked end-stage presence of fibrofatty tissue was seen in the post-mortem sections from Patient II.1 Family 6 (C). No mitochondrial abnormalities (D), rods or other inclusions were present in the muscle sample from affected foetus II.2 from Family 4 (D and E). Rods or other inclusions were also absent in Patient II.2 from Family 5 (F). Haematoxylin and eosin staining in the biopsies from 'classical' congenital myopathy cases showed myopathic features with abnormal fibre size variation without necrosis and regeneration (G–I). Mild fibrofatty replacement was present in the quadriceps biopsy taken from Patient II.1 Family 1 (age 2 years) (G), Patient II.1 Family 4 (age 1 month) (H) and Patient II.1 Family 3 (age 1.5 months) (I). NADH oxidative enzyme staining showed a population of small type I fibres with slow myosin in the biopsy of Patient II.1 Family 1 cohort member (J). This was confirmed with myosin heavy chain immunolabelling [fast myosin (M); slow myosin (N)]. Foetal myosin (O) showed scattered, abnormal, very small fibres measuring $<5\ \mu\text{m}$. NADH histochemistry showed preserved fibre typing without cores or minicores in Patient II.1 Family 4 (K). Rods or other inclusions were absent in the Gomori Trichrome stain (Patient II.1 Family 3) (L).

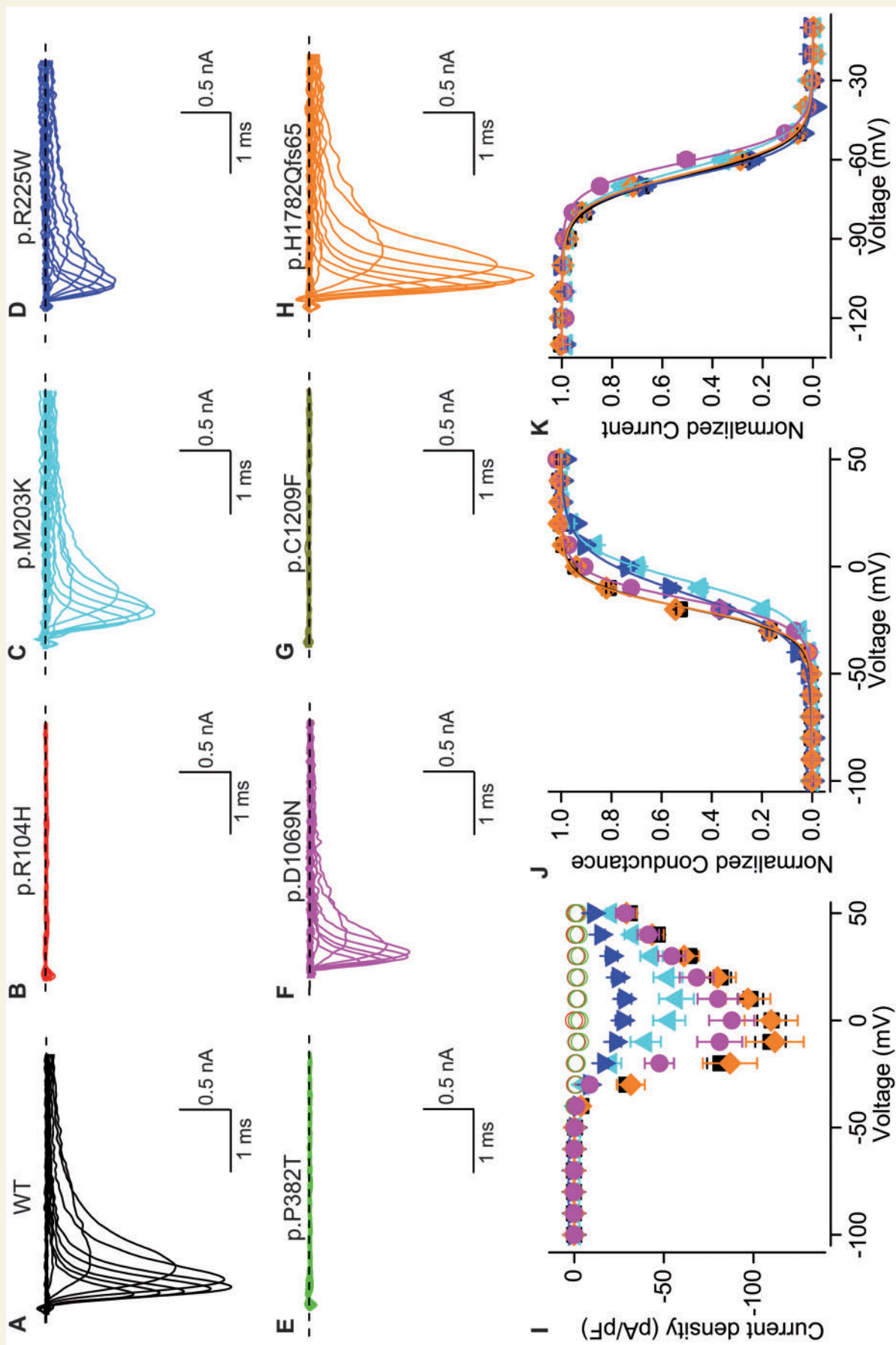


Figure 5 Electrophysiological characterization of congenital myopathy channel variants. Current traces in response to depolarizing test voltages for wild-type (WT; **A**), p.R104H (**B**), p.M203K (**C**), p.R225W (**D**), p.P382T (**E**), p.D1069N (**F**), p.C1209F (**G**) and p.H1782Qfs65 (**H**) channels expressed in HEK293 cells. Scale-bars are 1 ms (x-axis), 0.5 nA (y-axis). The dashed line represents the baseline current level at holding voltage. Mean current density (**I**) and normalized conductance (**J**) response to test voltages for channels expressed in HEK293 cells. Solid lines represent fit of Boltzmann equation to the mean data. Variants are colour coded as in **A–H**. Symbols: wild-type (square), p.M203K (triangle), p.R225W (inverted triangle), p.D1069N (circle), p.H1782Qfs65 (diamond), p.R104H, p.P382T and p.C1209F are all represented by open circles. Mean normalized currents in response to test voltage of -10 mV (HEK293) following a prepulse steps of 150 ms to voltages indicated in x-axis (**K**). Lines represent fit of Boltzmann equation to the mean data. Colour coding and symbols are as in **I**.

Table 3 Biophysical parameters of Na_v1.4 variants

Clone	Activation				Fast inactivation		
	V _{1/2} (mV)	V _{slope} (mV)	I _{peak} (at 0mV) (pA/pF)	n	V _{1/2} (mV)	V _{slope} (mV)	n
Wild-type (HEK293)	-20.0 ± 0.4	6.36 ± 0.12	-110.1 ± 8.0	51	-65.8 ± 0.4	5.55 ± 0.09	52
R104H	NA	NA	–	15	NA	NA	–
M203K	-7.0 ± 1.2***	8.47 ± 0.25***	-53.0 ± 9.0***	12	-63.1 ± 0.9*	5.98 ± 0.46	13
R225W	-12.3 ± 1.1***	10.68 ± 0.34***	-27.1 ± 4.1***	9	-66.0 ± 0.6	5.23 ± 0.28	13
P382T	NA	NA	–	13	NA	NA	–
D1069N	-15.8 ± 0.7***	6.24 ± 0.28	-87.8 ± 12.6	13	-60.2 ± 0.8***	5.15 ± 0.22	15
C1209F	NA	NA	–	11	NA	NA	–
H1782Qfs65	-20.2 ± 0.8	6.11 ± 0.22	-109.9 ± 14.8	16	-64.8 ± 0.4	5.62 ± 0.14	19

Mean ± SEM are shown. Statistical comparison of the variants was done against the wild-type channel using Student's *t*-test. Statistical significance is denoted by **P* < 0.05, ***P* < 0.01, ****P* < 0.001.

V_{1/2} = mid-point voltage; V_{slope} = slope factor; I_{peak} = peak current density at 0 mV; NA = not available.

In vitro electrophysiological characterization of the SCN4A mutations

Three of the variants (p.R104H, p.P382T and p.C1209F) did not display any detectable sodium currents when expressed in HEK293 cells (*n* > 10) (Fig. 5), suggesting that they render the channel completely non-functional.

Variants p.M203K, p.R225W and p.D1069N displayed currents with reduced amplitude (*P* < 0.001 for M203K and R225W, *P* = 0.07 for D1069N). The voltage dependence of channel activation for these three variants was shifted to depolarizing direction (*P* < 0.001) (Fig. 5 and Table 3). The data clearly demonstrate that the function of these channels is impaired. The loss-of-function effects of the p.D1069N mutation were confirmed in an alternate heterologous expression system, Chinese hamster ovary (CHO) cells (data not shown).

The frameshift variant p.H1782Qfs65 did not display any loss of Na_v1.4 channel function properties upon heterologous expression. Neither the peak current density, the voltage dependencies of the channel activation, fast- or slow inactivation, nor the time courses of onset or recovery from fast inactivation were significantly different from wild-type channels (Fig. 5, Table 3, Supplementary Fig. 2 and Supplementary Table 4).

In addition to impaired channel activation, the voltage dependence of fast inactivation was shifted to depolarizing direction for p.D1069N (*P* < 0.001) and p.M203K (*P* < 0.05) variants (Fig. 5 and Table 3). Some substitutions also displayed a statistically significant change in the voltage dependence of slow inactivation or in the kinetics of fast inactivation. Analysis of these data is shown in Supplementary Fig. 2 and Supplementary Table 4.

The p.R1135C channel has been comprehensively characterized (Groome *et al.*, 2014). It causes loss of channel function by enhancing fast inactivation of the channel. In addition it causes gating pore currents, which are typical for hypokalaemic periodic paralysis mutant channel. We

studied the gating pore currents of the p.R1135C channels using *X. laevis* oocyte expression system. We found that the gating pore current of the rat Na_v1.4-R1128C orthologue is active at hyperpolarized resting voltages even when the holding voltage is hyperpolarized (Supplementary Fig. 3). This differs from previous studies (Groome *et al.*, 2014) that detected the hyperpolarization activated gating pore currents only after a depolarizing prepulse.

Discussion

Using whole exome sequencing, we ascertained a cohort of 11 individuals from six unrelated and ethnically distinct kindreds with homozygous or compound heterozygous mutations in *SCN4A*. Affected individuals presented with congenital myopathy of variable severity. All had *in utero*- or neonatal-onset muscle weakness resulting in reduced foetal movements. Patients at the severe end of the spectrum had clinical features associated with foetal hypokinesia (maternal polyhydramnios, *in utero* upper/lower limb contractures, talipes and hydrops) resulting in intrauterine or early postnatal death. The patients at the less severe end of the spectrum presented with generalized hypotonia and weakness at birth or within the first few days of life. This was typically associated with mild-to-moderate facial muscle weakness without ptosis, and significant early respiratory and feeding difficulties. Surviving individuals had delayed motor milestones and achieved independent ambulation before the age of 3 years. Consistent with many forms of 'classical' congenital myopathy, all cohort members who survived infancy experienced improvement in their strength and motor skills over time, and concomitant resolution of early respiratory and feeding difficulties.

The phenotype of the cohort members described in this study differs from that of other sodium channelopathies (e.g. autosomal dominant myotonia and autosomal dominant periodic paralysis), and from the two reported *SCN4A*-related congenital myasthenia cases. In stark contrast to

other sodium channelopathies, the congenital myopathy cohort members had *in utero*- or neonatal-onset permanent muscle weakness, rather than later-onset episodic muscle weakness. Although individuals with periodic paralysis and, in rare instances, paramyotonia congenita, sometimes develop permanent proximal muscle weakness (Suetterlin *et al.*, 2014), this is typically a late complication of the condition and not a presenting feature. In addition, none of the patients had the typical features of weakness aggravated by specific factors (cold, special food and exercise) that trigger a worsening of stiffness and/or weakness in SCN4A-related myotonia or periodic paralysis (Cannon, 2015). While in one teenaged patient excessive muscle activity was associated with marked increase in fatigue, detailed electrophysiology examination failed to demonstrate neuromuscular junction defects or aspects evocative of periodic paralysis.

Other characteristic features of our patient cohort were the association with abnormal muscle histopathological features, and a selective pattern of muscle involvement on muscle MRI.

The majority of SCN4A mutations associated with periodic paralysis and myotonia are dominantly inherited and cause gain-of-function predominantly by enhancing channel activation, attenuating fast inactivation or by inducing gating pore currents. Mild enhancement of fast inactivation leading to a loss-of-function effect has been reported for some Na_v1.4 mutations in hypokalaemic periodic paralysis. Thus far, there are only two reports of loss of Na_v1.4 channel function and recessive inheritance of SCN4A mutations in congenital myasthenic syndrome (Tsuji *et al.*, 2003; Arnold *et al.*, 2015). The loss-of-function effects were caused by enhanced fast inactivation in both cases.

Recessive loss-of-function Na_v1.4 mutations as a pathomechanism of congenital myopathy

In this study, we establish that recessive SCN4A loss-of-function mutations are the underlying pathomechanism in patients affected by a novel form of congenital myopathy.

All 12 parents and one unaffected sibling were confirmed to be carriers of one SCN4A mutation but were unaffected clinically, based on formal examination in nine cases, and on history and verbal report in the other four. The absence of clinical findings in confirmed carriers suggests that the loss of function in only one allele is insufficient to cause a clinical phenotype, and further suggest an autosomal recessive mechanism of inheritance.

The inheritance of full loss-of-function mutations on both alleles appears to cause a particularly severe phenotype resulting in early lethality, evidenced by the homozygous p.P382T mutation detected in the three affected individuals from Family 5. This suggests that muscles without functional Na_v1.4 channels are unable to support life.

The remaining cohort members carried compound heterozygous SCN4A mutations. All had one mutation that resulted in a completely non-functional channel, either as a consequence of Na_v1.4 protein truncation or as a result of a mutation that led to a complete abolition of the sodium flux through the channel (p.R104H, p.C1209F, p.Q470X, p.A1049VfsX50, p.Y1593X in Families 1–4 and 6, respectively). Affected individuals from four of the five families with compound heterozygous mutations had a second mutation, in the *trans* allele, which resulted in partial Na_v1.4 loss-of-function mainly by attenuating channel activation (p.R225W, p.D1069N and p.M203K, Families 2, 4 and 6, respectively) or by enhancing channel inactivation (p.R1135C, Family 1). In Family 3, there were no detectable loss-of-function properties in HEK293 cells for one of the Na_v1.4 variants (discussed below).

Some of the affected individuals with compound heterozygous mutations (Families 6 and 4) also died *in utero*, or at birth, implying that some combinations of complete loss-of-function/partial loss-of-function mutations also result in a severe clinical phenotype. Interfamilial variability was, however, also clearly evident. This is best demonstrated in Family 4, where one affected infant developed severe *in utero*-onset hypokinesia and died within a few hours of birth. The other affected infant survived infancy albeit with significant feeding, and respiratory support. She is now 8 years of age, has a phenotype consistent with ‘classical’ congenital myopathy and continues to improve clinically over time, a feature often associated with ‘classical’ congenital myopathies. Further studies in a large cohort of patients are required to delineate more precise correlation of functional and clinical expression of SCN4A mutations.

One of the mutations present in the 2.5-year-old male cohort member from Family 3 (p.H1782Qfs65) did not result in loss of channel function when studied in HEK293 cells. This child has a similar clinical phenotype to other cohort members. He was born following a pregnancy complicated by polyhydramnios (first noted at delivery), had marked neonatal-onset weakness and hypotonia associated with facial weakness, required significant feeding and respiratory support from birth, but continues to improve clinically. The frameshift mutation is predicted to alter the C-terminal of the protein and extends it by nine residues compared to the wild-type channel. Our analysis suggests that in HEK293 cells, Na_v1.4 channel C-terminus has no detectable effects on the biophysical properties of the channel. This is in agreement with previous data showing that a deletion of the C-terminal 100 residues has no effect on channel gating (Herzog *et al.*, 2003). However, it is possible that the frameshift mutation disrupts channel function in a tissue-specific manner, which cannot be detected in a heterologous expression system, or it might cause loss-of-function effect not elicited by the functional analyses undertaken in this study. For example, the last five C-terminal amino acid residues play an important role in the interaction of Na_v1.4 with the PDZ domain of syntrophin (Schultz *et al.*, 1998), a scaffold protein that is a part

of the dystrophin-associated protein complex and has a vital role in the maintenance of muscle integrity (Ehmsen *et al.*, 2002). Alterations of the sequence of the C-terminus may also disrupt signals responsible for channel trafficking, targeting, activity or stability. Additional studies are required to further investigate these possibilities.

Interestingly, we identified compound heterozygosity for p.R1135C and a full loss-of-function mutation in a 14-year-old girl with a ‘classical’ congenital myopathy phenotype. Homozygous p.R1135C has been previously described in a patient with an unusual recessive form of hypokalaemic periodic paralysis (Groome *et al.*, 2014). Consistent for periodic paralysis, the mutant channels display gating pore currents. The mutation also causes loss-of-function effects on the main pore current by enhancing fast inactivation, which results in reduced channel availability. However, the heterozygous carrier of this mutation in our family had no periodic weakness. Furthermore the muscle weakness of the proband, carrying a heterozygous p.R1135C and a full loss-of-function *SCN4A* mutation in trans, has been permanent rather than periodic, suggesting that the reduced main pore sodium current is the likely pathomechanism of the myopathy. Although this patient, who is now 14 years old, has started to complain of recurrent brief episodes of increased muscle weakness, a detailed electrophysiological study failed to identify any abnormalities typical of periodic paralysis, myotonia or neuromuscular junction abnormalities, but only demonstrated myopathic features.

Our findings are in agreement with Na_v1.4 being the main skeletal muscle voltage gated sodium channel isoform, including the foetal and neonatal muscle, critical to muscle function (Zhou and Hoffman, 1994). Expression of a second isoform, Na_v1.5, which is tetrodotoxin resistant, has been shown in developing and denervated muscle (Harris and Thesleff, 1971; Weiss and Horn, 1986). The tetrodotoxin resistant sodium current may be able to generate action potentials in muscle of newborn rats (Harris and Marshall, 1973). However, these action potentials are defective and disappear soon after birth, confirming a key role for tetrodotoxin sensitive currents in generating skeletal muscle action potentials. Possible differences in Na_v1.5 expression in foetal muscle may, however, contribute to the variable clinical presentation in the myopathy cohort. No mutations in our cohort were detected in *SCN5A* that encodes Na_v1.5, the principal sodium channel responsible for the initiation of the cardiac action potential (George *et al.*, 1995). This is consistent with the lack of cardiac involvement in the cohort members.

In conclusion, we have identified homozygous or compound heterozygous recessive mutations in *SCN4A* in six congenital myopathy families. At least one of the mutations in each patient renders the Na_v1.4 channel fully non-functional. The mutation on the second allele resulted in full or partial loss of Na_v1.4 channel function in five out of six families. This constitutes the first report describing full

loss-of-function *SCN4A* mutations as associated with a clinical phenotype.

The Na_v1.4 channel is crucial for the initiation and propagation of the muscle action potential. The link between the depolarization of the membrane and the Ca²⁺ release triggering the muscle contraction is mediated by the interaction between the voltage-sensing DHPR and the Ca²⁺ release channel RYR1, main components of the excitation–contraction coupling. Mutations in *RYR1* are an established cause of several types of congenital myopathy (Zhang *et al.*, 1993; Jungbluth *et al.*, 2002; Monnier *et al.*, 2003). Here, we propose that the combined effects of two loss-of-function mutant Na_v1.4 channels, one of which is a full loss-of-function, attenuates the action potential amplitude in skeletal muscle to a level insufficient to sustain normal muscle force.

These findings have important implications for genetic diagnosis as they confirm a new form of recessive *SCN4A*-related congenital myopathy, a discovery which will undoubtedly result in improved genetic diagnosis rates in patients with this group of disorders. Furthermore, this study indicates that *SCN4A* is another example of a growing number of genes encoding for ion channels recently found to be involved in congenital myopathies. Our study also opens research avenues aimed at improving the therapeutic management of these patients.

Acknowledgements

We gratefully acknowledge the clinical and scientific support from our colleagues Dr Mariacristina Scoto and Darren Chambers from the Dubowitz Neuromuscular Centre, UCL Institute of Child Health, London and Dr W. Wasserman, Dr M. Tarailo-Graovac, Dr C.J. Ross, Dr H. Gill, Dr K. Selby, Dr B. Sayson, Dr P Eydoux, Ms M. Balicki, Ms Chieko Chijiwa from the University of British Columbia, Vancouver CA.

Funding

The research leading to these results has received funding from the European Community’s Seventh Framework Programme (FP7/2007-2013) under grant agreement n° 2012-305121 ‘Integrated European –omics research project for diagnosis and therapy in rare neuromuscular and neurodegenerative diseases (NEUROMICS)’. This study was also supported by the National Institute for Health Research Biomedical Research Centre at Great Ormond Street Hospital for Children NHS Foundation Trust and University College London (F.M.). The support of the Medical Research Council Neuromuscular Centre Biobank and of the Muscular Dystrophy Campaign study coordinators is also gratefully acknowledged (F.M.; M.G.H.). M.G.T. was funded by a Medical Research Council Centre for Neuromuscular Diseases studentship.

R.M. was funded by United Kingdom Medical Research Council project grant MR/M006948/1. We also acknowledge support of the UCLH Biomedical Research Centre. E.C.O. was gratefully funded by a Winston Churchill Memorial Trust of Australia fellowship and by a National Health and Medical Research (NHMRC) early career research fellowship (grant number: GNT1090428). This research was also supported by the National Health and Medical Research Council of Australia (Early Career Researcher Fellowship #1035955 to G.R., Research Fellowship APP1002147 to N.G.L. and Project Grant APP1022707; EU Collaborative grant APP1055295); the Association Francaise contre les Myopathies (#15734). We acknowledge funding support from the B.C. Children's Hospital Foundation (Treatable Intellectual disability Endeavour in British Columbia: 1st Collaborative Area of Innovation www.tidebc.org), Genome BC (SOF-195) and Canadian Institutes of Health Research (#301221). Dr. C. van Karnebeek is a recipient of the Michael Smith Foundation for Health Research Scholar Award. M.G.H. and R.M. work is supported by the UCLH Biomedical Research Centre.

Supplementary material

Supplementary material is available at *Brain* online.

References

- Arnold WD, Feldman D, Ramirez S, He L, Kassar D, Quick A, et al. Defective fast inactivation recovery of Nav1.4 in congenital myasthenic syndrome. *Ann Neurol* 2015; 77: 840–50.
- Cannon SC. Channelopathies of skeletal muscle excitability [Review]. *Compr Physiol* 2015; 5: 761–90.
- Corrochano S, Männikkö R, Joyce PI, McGoldrick P, Wettstein J, Lassi G, et al. Novel mutations in human and mouse SCN4A implicate AMPK in myotonia and periodic paralysis. *Brain* 2014; 137: 3171–85.
- de Ligt J, Willemsen MH, van Bon BW, Kleefstra T, Yntema HG, Kroes T, et al. Diagnostic exome sequencing in persons with severe intellectual disability. *N Engl J Med* 2012; 367: 1921–9.
- Ehmsen J, Poon E, Davies K. The dystrophin-associated protein complex [Review]. *J Cell Sci* 2002; 115: 2801–3.
- George AL Jr, Varkony TA, Drabkin HA, Han J, Knops JF, Finley WH, et al. Assignment of the human heart tetrodotoxin-resistant voltage-gated Na⁺ channel alpha-subunit gene (SCN5A) to band 3p21. *Cytogenet Cell Genet* 1995; 68: 67–70.
- Groome JR, Lehmann-Horn F, Fan C, Wolf M, Winston V, Merlini L, et al. Nav1.4 mutations cause hypokalaemic periodic paralysis by disrupting IIIIS4 movement during recovery. *Brain* 2014; 137: 998–1008.
- Harris JB, Marshall MW. Tetrodotoxin-resistant action potentials in newborn rat muscles. *Nat New Biol* 1973; 243: 191–2.
- Harris JB, Thesleff S. Studies on tetrodotoxin dependent action potentials in denervated skeletal muscle. *Acta Physiol Scand* 1971; 83: 382–8.
- Herzog RI, Liu C, Waxman SG, Cummins TR. Calmodulin binds to the C terminus of sodium channels Nav1.4 and Nav1.6 and differentially modulates their functional properties. *J Neurosci* 2003; 23: 8261–70.
- Jungbluth H, Müller CR, Halliger-Keller B, Brockington M, Brown SC, Feng L, et al. Autosomal recessive inheritance of RYR1 mutations in a congenital myopathy with cores. *Neurology* 2002; 59: 284–7.
- Lee SC, Kim HS, Park YE, Choi YC, Park KH, Kim DS. Clinical diversity of SCN4A-mutation-associated skeletal muscle sodium channelopathy. *J Clin Neurol* 2009; 5: 186–91.
- Matthews E, Guet A, Mayer M, Vicart S, Pemble S, Sternberg D, et al. Neonatal hypotonia can be a sodium channelopathy: recognition of a new phenotype. *Neurology* 2008; 71: 1740–2.
- Monnier N, Ferreira A, Marty I, Labarre-Vila A, Mezin P, Lunardi J. A homozygous splicing mutation causing a depletion of skeletal muscle RYR1 is associated with multi-minicore disease congenital myopathy with ophthalmoplegia. *Hum Mol Genet* 2003; 12: 1171–8.
- Nance JR, Dowling JJ, Gibbs EM, Bönnemann CG. Congenital myopathies: an update [Review]. *Curr Neurol Neurosci Rep* 2012; 12: 165–74.
- Ravenscroft G, Laing NG, Bönnemann CG. Pathophysiological concepts in the congenital myopathies: blurring the boundaries, sharpening the focus [Review]. *Brain* 2015; 138: 246–68.
- Ravenscroft G, Sollis E, Charles AK, North KN, Baynam G, Laing NG. Fetal akinesia: review of the genetics of the neuromuscular causes [Review]. *J Med Genet* 2011; 48: 793–801.
- Ryan AM, Matthews E, Hanna MG. Skeletal-muscle channelopathies: periodic paralysis and nondystrophic myotonias [Review]. *Curr Opin Neurol* 2007; 20: 558–63.
- Schultz J, Hoffmüller U, Krause G, Ashurst J, Macias MJ, Schmieder P, et al. Specific interactions between the syntrophin PDZ domain and voltage-gated sodium channels. *Nat Struct Biol* 1998; 5: 19–24.
- Sewry CA, Jimenez-Mallebrera C, Muntoni F. Congenital myopathies [Review]. *Curr Opin Neurol* 2008; 21: 569–75.
- Stühmer W, Conti F, Suzuki H, Wang XD, Noda M, Yahagi N, et al. Structural parts involved in activation and inactivation of the sodium channel. *Nature* 1989; 339: 597–603.
- Suetterlin K, Männikkö R, Hanna MG. Muscle channelopathies: recent advances in genetics, pathophysiology and therapy [Review]. *Curr Opin Neurol* 2014; 27: 583–90.
- Trimmer JS, Cooperman SS, Tomiko SA, Zhou JY, Crean SM, Boyle MB, et al. Primary structure and functional expression of a mammalian skeletal muscle sodium channel. *Neuron* 1989; 3: 33–49.
- Tsujino A, Maertens C, Ohno K, Shen XM, Fukuda T, Harper CM, et al. Myasthenic syndrome caused by mutation of the SCN4A sodium channel. *Proc Natl Acad Sci USA* 2003; 100: 7377–82.
- Vicart S, Sternberg D, Fontaine B, Meola G. Human skeletal muscle sodium channelopathies [Review]. *Neurol Sci* 2005; 26: 194–202.
- Wallgren-Petersson C and Laing N The congenital myopathies. In: Karpati G, Hilton-Jones D, Bushby K, Griggs RC, editors. *Disorders of voluntary muscle*. Cambridge: Cambridge University Press; 2010. pp. 282–98.
- Weiss RE, Horn R. Functional differences between two classes of sodium channels in developing rat skeletal muscle. *Science* 1986; 233: 361–4.
- Zhang Y, Chen HS, Khanna VK, De Leon S, Phillips MS, Schappert K, et al. A mutation in the human ryanodine receptor gene associated with central core disease. *Nat Genet* 1993; 5: 46–50.
- Zhou J, Hoffman EP. Pathophysiology of sodium channelopathies. Studies of sodium channel expression by quantitative multiplex fluorescence polymerase chain reaction. *J Biol Chem* 1994; 269: 18563–71.